

CERTIFICATION

AOAC Research Institute Performance Tested MethodsSM

Certificate No. 010404

The AOAC Research Institute hereby certifies the method known as:

Compact Dry "Nissui" Total Count

manufactured by NISSUI Pharmaceutical Co., Ltd. 3-24-6, Ueno Taito-ku, Tokyo Japan 110-8736

This method has been evaluated in the AOAC Research Institute *Performance Tested Methods*SM Program and found to perform as stated in the applicability of the method. This certificate indicates an AOAC Research Institute Certification Mark License Agreement has been executed which authorizes the manufacturer to display the AOAC Research Institute *Performance Tested Methods*SM certification mark on the above-mentioned method for the period below. Renewal may be granted by the Expiration Date under the rules stated in the licensing agreement.

Scott Coates

Scott Coates, Senior Director Signature for AOAC Research Institute Issue Date Expiration Date January 9, 2023 December 31, 2023

2275 Research Blvd., Ste. 300, Rockville, Maryland, USA Telephone: +1-301-924-7077 Fax: +1-301-924-7089 Internet e-mail: aoacri@aoac.org * World Wide Web Site: http://www.aoac.org

AUTHORS ORIGINAL VALIDATION: Hidema Hajime Teramura, and Tadanobu MODIFICATION APRIL 2015: Shir MODIFICATION DECEMBER 2020	ı Nirazuka ngo Mizuochi	SUBMITTING COMPANY NISSUI Pharmaceutical Co., Ltd. 3-24-6, Ueno Taito-ku, Tokyo Japan 110-8736
METHOD NAME		CATALOG NUMBERS
Compact Dry "Nissui" Total Count	t	Nissui 06740 (40 plates), 06741 (240 plates)
INDEPENDENT LABORATORY ORIGINAL VALIDATION National Public Health Service for Wales Llandough Hospital Penlan Rd, Penarth, United Kingdom	MODIFICATION APRIL 2015, DECEMBER 2020 Campden BRI Station Road Chipping Campden Gloucerstershire, GL55 6LD, UK	EXPERT REVIEWERS ORIGINAL VALIDATION: AOAC RI PTM reviewers Yi Chen ¹ , Yvonne Salfinger ² , Maria Christina Fernandez ³ ¹ Food and Drug Administration, Center for Food Safety and Applied Nutrition, Maryland, USA ² Consultant, Colorado, USA ³ University of Buenos Aires, Buenos Aires, ARGENTINA
APPLICABILITY OF METHOD Target Organism – Aerobic micro Matrixes – raw meat (50 g) (raw lamb, raw veal, raw ground beef MODIFICATION APRIL 2015 – coo lettuce (10 g), milk powder (10 g pasteurized milk (1 mL) MODIFICATION DECEMBER 2020 chicken breast (50 g) Performance claims – The study difference between the Compact method and the reference method	ground pork, raw pork, raw) oked chicken (10g),), frozen fish (10g), 1 – raw ground beef (50g), raw data detected no statistical t Dry "Nissui" Total Count	REFERENCE METHODS Official Methods of Analysis 19 th Ed., AOAC INTERNATIONAL, Rockville, MD, Method 966.23, Microbiological Methods (2) ISO 4ISO 4833:2003, Microbiology of food and animal feeding stuffs – Horizontal Method for the enumeration of microorganisms – Colony-count technique at 30°C. (3) FSIS MLG 3.02 Quantitative Analysis of Bacteria in Foods as Sanitary Indicators: Chapter 3.6 Aerobic Plate Count (6)
ORIGINAL CERTIFICATION DATE	E	CERTIFICATION RENEWAL RECORD
February 2004		Renewed annually through December 2023.
METHOD MODIFICATION RECO 1. April 2015 Level 2 2. February 2019 Level 3. December 2020 Level	2	 SUMMARY OF MODIFICATION Matrix extension to include cooked chicken, pre-washed bagged shredded iceberg lettuce, frozen cod fillets, instant non-fat dry milk, pasteurized milk (2%). Shelf life increased to 24 months and corporate address change. Matrix extension for raw chicken breast enumeration at 24 ± 2 h and 48 ± 3 h. Added claim for raw ground beef at 24 ± 2 h.
Under this AOAC <i>Performance</i> 010404 this method is distribut 1. Hardy Diagnostics 2. R-Biopharm AG	<i>Tested MethodssM</i> License Number, ted by:	Under this AOAC <i>Performance Tested Methods</i> ^{5M} License Number, 010404 this method is distributed as: 1. Compact Dry TC 2. Compact Dry TC

PRINCIPLE OF THE METHOD (1)

Compact Dry TC qualifies as a rapid method kit for determining aerobic colony counts in foods. The plates are presterilized and contain culture medium and a cold-soluble gelling agent. The medium is rehydrated by inoculating 1 mL diluted sample into the center of the self-diffusible medium and allowing the solution to diffuse by capillary action. The plates can then be incubated and the colonies counted without any additional steps. The Compact Dry TC method was validated with 5 different raw meats. The performance tests were conducted at 35° and 30°C. In all required performance studies, no apparent differences were observed between the Compact Dry TC method and the Standard Pour Plate method (AOAC Official Method **966.23**) for the detection level of aerobic microorganisms. For the accuracy claim (n = 60), a correlation factor of $r^2 35 = 0.9977$ (35°C) and $r^2 30 = 0.9932$ (30°C) could be assigned, as stated in the application for "Performance Tested MethodSM" Quality consistency and storage robustness studies, showed no significant variations in plate count results with different production lots or plates of diverse storage age.

DISCUSSION OF THE VALIDATION STUDY (1)

Tal

The retested results, as instructed by AOAC RI with exactly the same sample preparation procedure as AOAC Official Method **966.23** showed equal results on both the Compact Dry TC Method and AOAC Official Method **966.23**. As evidenced by attached data of 5 replicates for 3 levels (4 levels for raw veal) of 5 kinds of raw meat (n = 80) through internal and independent validation study, r^2 as correlation coefficient for both methods was 0.9965. This means that the retested results are the same as the original, and therefore discussion on the original report must be valid.

Regarding the measured value, for only the lowest level, the value between Compact Dry TC and AOAC Official Method **966.23** differed from each other more than the range of the standard deviation, and by one-way ANOVA(*P*< 0.05) in independent validation studies. This phenomenon could not be observed with the other bacterial load levels and with the other 4 matrixes (raw ground pork, raw pork, raw lamb, and raw veal) by internal validation study. For all remaining levels and samples by internal validation study, the standard deviation of analogous samples delivered comparable values for the Compact Dry TC method and the AOAC Official Method **966.23**. Consequently, the results from the Compact Dry TC method for the repeatability study correspond with the results from pour plate methods.

Using an incubation temperature of 30°C instead of the recommended 35°C, which is the more common incubation temperature for total aerobic count estimations in the food regulations of the European Community, the results of the repeatability, accuracy, lot consistency, and lot stability obtained from the different matrixes were similar to those at 35°C. Using yogurt in the repeatability study, the 30°C incubation series exhibited major differences. The counts of the dilutions 104 and 103 measured with Compact

Dry TC were significantly lower than those with the pour plate method. Dilution 2.5×10^2 and 10^2 did not give any count with the Compact Dry TC technique. Because those results with yogurt could not be observed in the accuracy, lot consistency, and lot stability study at 30°C incubation, the results of the repeatability study might be attributed to internal errors.

The big advantages of the Compact Dry TC system are the reduced hands-on time and economical usage, as confirmed by the independent laboratory. In terms of plate preparation, inoculation, and reading the result, the Compact Dry TC system was easier and quicker than the conventional pour plate technique. Reading the plates was faster with the Compact Dry TC system, with the TTC indicator speeding up counting. It was observed that food particles, when present, did not appear to absorb the indicator. Food particles in the pour plate made reading plates and counting colonies relatively more difficult. For the Compact Dry TC system, less training is required than for the pour plate technique. Instructions on the use of the Compact Dry TC are clear and unambiguous. The Compact Dry TC system would also bring advantages in reduced storage space, waste disposal, and required incubator space. The long shelf life of the product also has benefits compared to ready-prepared agar, which has a limited shelf life and therefore requires more logistical planning.

			Compact Dry TCa	aerobic bacteria	AOAC 966.23 a	erobic bacteria
Level	Spike, CFU/g	Replicate	CFU/g	Log ₁₀ CFU/g	CFU/g	log ₁₀ CFU/
About 10 ² CFU/gLot 1	0°	1	525	2.72	500	2.70
		2	680	2.83	670	2.83
		3	430	2.63	415	2.62
		4	635	2.80	635	2.80
		5	365	2.56	355	2.55
		Mean	527	2.71	515	2.70
		Sr	133	0.11	136	0.12
		RSD _r , %	25.2	4.18	26.5	4.37
About 10 ⁴ CFU/gLot 2	0ª	1	78000	4.89	70500	4.85
		2	80000	4.90	76000	4.88
		3	77000	4.89	78000	4.89
		4	99000	5.00	96000	4.98
		5	55000	4.74	42500	4.63
		Mean	77800	4.88	72600	4.85
		Sr	15600	0.09	19400	0.13
		RSD _r , %	20.1	1.87	26.7	2.72
About 10 ⁶ CFU/gLot 3	0°	1	6400000	6.81	5850000	6.77
		2	9100000	6.96	8850000	6.95
		3	9350000	6.97	9150000	6.96
		4	6800000	6.83	6400000	6.81
		5	9300000	6.97	8250000	6.92
		Mean	8190000	6.91	7700000	6.88
		sr	1460000	0.08	1490000	0.09
		RSD _r , %	17.8	1.17	19.3	1.27

^a Naturally contaminated samples.

			Compact Dry TCa	aerobic bacteria	AOAC 966.23 ae	robic bacteria
Level	Spike, CFU/g	Replicate	CFU/g	Log ₁₀ CFU/g	CFU/g	Log ₁₀ CFU/g
About 10 ² CFU/gLot 1	0 <i>°</i>	1				
			125	2.10	165	2.22
		2	180	2.26	170	2.23
		3	100	2.00	90	1.95
		4	145	2.16	100	2.00
		5	120	2.08	170	2.23
		Mean	134	2.12	139	2.13
		Sr	30.3	0.10	40.4	0.14
		RSD _r , %	22.6	4.52	29.0	6.46
About 10 ⁴ CFU/gLot 2	0 ^{<i>a</i>}	1	37500	4.57	37500	4.57
		2	35000	4.54	35000	4.54
		3	96500	4.98	96500	4.98
		4	16100	4.21	15800	4.20
		5	18350	4.26	10400	4.02
		Mean	40690	4.51	39040	4.46
		Sr	32600	0.31	34200	0.37
		RSD _r , %	80.2	6.85	87.6	8.38
About 10 ⁶ CFU/gLot 3	0 <i>°</i>	1	3700000	6.57	3500000	6.54
		2	2855000	6.46	2840000	6.45
		3	7800000	6.89	7550000	6.88
		4	1210000	6.08	1300000	6.11
		5	1545000	6.19	1775000	6.25
		Mean	3420000	6.44	3393000	6.45
		Sr	2640000	0.32	2480000	0.29
		RSD _r , %	77.3	4.98	73.1	4.56

^a Naturally contaminated samples.

			Compact Dry TCa	aerobic bacteria	AOAC 966.23 aerob	ic bacteria
Level	Spike, CFU/g	Replicate	CFU/g	Log ₁₀ CFU/g	CFU/g	Log ₁₀ CFU/
About 10 ² CFU/gLot 1	0 ^{<i>a</i>}	1				
			255	2.41	225	2.35
		2	370	2.57	360	2.56
		3	455	2.66	410	2.61
		4	145	2.16	130	2.11
		5	840	2.92	965	2.98
		Mean	413	2.54	418	2.52
		Sr	266	0.28	325	0.32
		RSD _r , %	64.4	11.2	77.8	12.8
About 10 ⁴ CFU/gLot 2	0 <i>°</i>	1	19450	4.29	18900	4.28
		2	13400	4.13	13600	4.13
		3	83500	4.92	79000	4.90
		4	15000	4.18	14200	4.15
		5	10300	4.01	9700	3.99
		Mean	28330	4.31	27100	4.29
		Sr	31000	0.36	29200	0.36
		RSD _r , %	109	8.33	108	8.28
About 10 ⁶ CFU/gLot 3	0 ^{<i>a</i>}	1	5850000	6.77	4800000	6.68
-		2	1610000	6.21	1120000	6.05
		3	2020000	6.31	2025000	6.31
		4	3700000	6.57	3500000	6.54
		5	1290000	6.11	1320000	6.12
		Mean	2890000	6.39	2553000	6.34
		Sr	1900000	0.27	1565118.206	0.27
		RSD _r , %	65.5	4.23	61.30506096	4.26

^a Naturally contaminated samples.

			Compact Dry TCae	erobic bacteria	AOAC 966.23 ae	robic bacteria
Level	Spike, CFU/g	Replicate	CFU/g	Log ₁₀ CFU/g	CFU/g	Log ₁₀ CFU/g
0 CFU/g Lot 1	None	1	5	0.70	5	0.70
		2	10	1.00	5	0.70
		3	20	1.30	10	1.00
		4	10	1.00	5	0.70
		5	10	1.00	10	1.00
		Mean	11	1.00	7	0.82
		sr	5.48	0.21	2.74	0.16
		RSD _r , %	49.8	21.3	39.1	20.1
About 10 ² CFU/g Lot 1	250	1	260	2.41	256	2.41
		2	224	2.35	238	2.38
		3	170	2.23	172	2.24
		4	292	2.47	284	2.45
		5	220	2.34	230	2.36
		Mean	233.2	2.36	236	2.37
		sr	45.9	0.09	41.4	0.08
		RSD _r , %	19.7	3.75	17.5	3.44
About 10⁴ CFU/g Lot 2	25000	1	23200	4.37	23200	4.37
		2	11800	4.07	11900	4.08
		3	13700	4.14	13580	4.13
		4	56000	4.75	53800	4.73
		5	14680	4.17	14800	4.17
		Mean	23900	4.30	23500	4.29
		sr	18500	0.27	17500	0.27
		RSD _r , %	77.4	6.39	74.7	6.21
About 10 ⁶ CFU/g Lot 3	1500000	1	1340000	6.13	1320000	6.12
		2	1780000	6.25	1540000	6.19
		3	860000	5.93	1210000	6.08
		4	1100000	6.04	1290000	6.11
		5	1540000	6.19	1340000	6.13
		Mean	1324000	6.11	1340000	6.13
		sr	361000	0.12	122000	0.04
		RSD _r , %	27.2	2.03	9.12	0.63

ethod comparison (indep			Compact Dry TCa	erobic bacteria	AOAC 966.23 a	erobic bacteria
Level	Spike, CFU/g	Replicate	CFU/g	Log ₁₀ CFU/g	CFU/g	Log ₁₀ CFU/
About 10⁴ CFU/gLot 1	0 ^{<i>a</i>}	1	15300	4.18	25400	4.40
		2	9600	3.98	24650	4.39
		3	10900	4.04	20200	4.31
		4	11100	4.05	27950	4.45
		5	10600	4.03	20700	4.32
		Mean	11500	4.06	23780	4.37
		sr	2200	0.08	3280	0.06
		RSD _r , %	19.1	1.89	13.8	1.38
About 10 ⁶ CFU/gLot 2	0 <i>°</i>	1	1590000	6.20	1300000	6.11
		2	1660000	6.22	1335000	6.13
		3	1870000	6.27	1780000	6.25
		4	1110000	6.05	980000	5.99
		5	1850000	6.27	1680000	6.23
		Mean	1616000	6.20	1415000	6.14
		sr	307000	0.09	321000	0.10
		RSD _r , %	19.0	1.49	22.7	1.68
About 10 ⁸ CFU/gLot 3	0 ^{<i>a</i>}	1	93000000	7.97	86500000	7.94
		2	125000000	8.10	111500000	8.05
		3	164000000	8.21	128000000	8.11
		4	12000000	8.08	10400000	8.02
		5	19000000	8.28	158500000	8.20
		Mean	138400000	8.13	117700000	8.06
		sr	38400000	0.12	27250000	0.10
		RSD _r , %	27.7	1.50	23.2	1.22

^a Naturally contaminated samples.

DISCUSSION OF MODIFICATION STUDY APPROVED APRIL 2015 (4)

For this matrix extension study, the Compact Dry TC was evaluated at two time points, 48 ± 3 h, which is recommended by Nissui and 72 ± 3 h, which is consistent with ISO 4833. Results from each time point were compared to ISO 4833 and to each other. Statistical analyses indicate that there were no significant differences in enumeration of total aerobic counts in cooked chicken, lettuce, frozen fish, milk powder and pasteurized milk between the two time points tested, and thus 48 h is still recommended.

In the single laboratory matrix studies for cooked chicken, lettuce, frozen fish, and milk powder, statistical differences were found between the Compact Dry TC and ISO 4833 for two levels each out of five levels tested for lettuce and frozen fish. In both matrixes, the differences were found in the mid and high contamination levels. For the 48 h Compact Dry TC time point, the mean log₁₀ differences were >0.5 (0.702 and 0.624 for lettuce, and 0.708 and 0.522 for frozen fish), and although the CIs were small [(0.601, 0.802) and (0.529, 0.719) for lettuce, and (0.538, 0.879) and (0.448, 0.595) for fish], the CIs were outside (-0.5, 0.5) acceptance criterion. In each case, the reference method recorded higher results than the Compact Dry TC. For the other three levels tested for lettuce and fish, mean differences were small (<0.5) and CIs were within (-0.5, 0.5). No significant differences were found in cooked chicken and milk powder. Statistical outliers were detected using Grubb's test in cooked chicken and lettuce, but no justifiable cause was noted in the study for removing the outliers, and so all data were included in the analysis. The r² was >0.97 for all matrixes.

In the multi-laboratory study on pasteurized milk, no statistical differences were found between any of the method comparisons after Laboratory 4 was removed from the analysis for unexpectedly high counts in the uncontaminated samples. Due to shipping and scheduling issues, four laboratories initiated testing one day later than the other ten laboratories. An initial review of the data indicated no significant differences in results between the two start dates, so all laboratories (with the exception of Laboratory 4) were included in the statistical analysis. Across 14 data sets, the mean differences between Compact Dry TC at 48 and 72 h were -0.001 to 0.001 ($r^2 = 1.0$), and the mean differences between the Compact Dry TC at 48 h and ISO 4833 were -0.60 to 0.101 ($r^2 = 0.98$). The CIs on the mean differences were well within the (-0.5, 0.5) acceptance criterion, and even smaller (less than -0.2, 0.2) for the contaminated samples. The s_r, RSD_r, s_R, and RSD_R were equally small. These data indicate that the Compact Dry TC gives consistent results, comparable to the reference method, across laboratories.

	N DATA APPROVE laboratory matri	· · · ·) t Dry TC 48 h vs IS	O 4833 (4)							
Matuiu	Cont. level —	Co	ompact Dry TC 48 h	ı		ISO 4833		Mean diff. ^d	95%	6 Cl ^e	- r ^{2h}
Matrix	Cont. level	Mean ^a	Sr ^b	RSD _r ^c	Mean	Sr	RSDr	iviean diff."	LCL ^f	UCL ^g	- r²"
	1	0.281	0.594	211	0.357	0.757	212	0.076	-0.070	0.219	
	2	2.399	0.304	12.7	2.572	0.517	20.1	0.173	-0.113	0.459	
Cooked chicken	3	3.309	0.364	11.0	3.383	0.357	10.6	0.074	-0.015	0.163	0.99
	4	5.056	0.284	5.62	5.404	0.133	2.46	0.348	0.207	0.490	
	5	6.173	0.252	4.08	6.194	0.209	3.37	0.021	-0.036	0.077	
	1	2.346	0.663	28.3	2.663	0.682	25.6	0.317	0.156	0.478	
	2	3.359	0.709	21.1	3.437	0.346	10.1	0.078	-0.230	0.387	
Lettuce	3	4.491	0.324	7.21	5.193	0.352	6.78	0.702	0.601	0.802	0.98
	4	5.443	0.201	3.69	5.638	0.244	4.28	0.196	0.129	0.263	
	5	7.010	0.143	2.04	7.634	0.134	1.76	0.624	0.529	0.719	
	1	2.953	0.178	6.03	3.089	0.158	5.11	0.136	0.016	0.256	
	2	3.924	0.121	3.08	4.052	0.091	2.25	0.128	0.056	0.200	
Frozen fish	3	4.791	0.191	3.99	5.499	0.139	2.53	0.708	0.538	0.879	0.97
	4	5.854	0.135	2.31	6.246	0.091	1.46	0.392	0.326	0.459	
	5	6.432	0.066	1.03	6.953	0.090	1.29	0.522	0.448	0.595	
	1	0.645	0.845	131	1.012	0.736	72.7	0.367	-0.067	0.800	
	2	2.064	0.139	6.73	2.232	0.157	7.03	0.168	0.082	0.253	
Milk powder	3	4.026	0.134	3.33	3.993	0.076	1.90	-0.033	-0.102	0.036	0.98
F 3 11 6 6 1	4	5.010	0.110	2.20	4.926	0.103	2.09	-0.084	-0.149	-0.018	
	5	5.709	0.132	2.31	5.639	0.171	3.03	-0.070	-0.127	-0.012	

^aMean of five replicate portions, plated in duplicate, after logarithmic transformation: Log₁₀[CFU/g + (0.1)f].

^bRepeatability standard deviation.

^cRelative standard deviation for repeatability.

^{*d*}Mean difference between the candidate and reference methods.

^eConfidence interval.

^f95% Lower confidence limit for difference of means.

^{*g*}95% Upper confidence limit for difference of means.

^hSquare of correlation coefficient.

Table 2. Single laboratory matrix study: Compact Dry TC 72 h vs ISO 4833 (4)

		Co	ompact Dry TC 72	h		ISO 4833		a a luce d	95%	S CI ^e	r ^{2h}
Matrix	Cont. level	Mean ^a	Sr ^b	RSD _r ^c	Mean	Sr	RSDr	– Mean diff. ^d –	LCL ^f	UCL ^g	-
	1	0.281	0.594	211	0.357	0.757	212	0.076	-0.068	0.219	
	2	2.734	0.636	23.4	2.572	0.517	20.1	-0.163	-0.358	0.033	
Cooked chicken	3	3.323	0.369	11.1	3.383	0.357	10.6	0.060	-0.033	0.153	0.99
	4	5.063	0.280	5.53	5.404	0.133	2.46	0.342	0.203	0.480	
	5	6.179	0.249	4.03	6.194	0.209	3.37	0.015	-0.040	0.070	
	1	2.496	0.593	23.8	2.663	0.682	25.6	0.168	0.040	0.296	
	2	3.393	0.732	21.6	3.437	0.346	10.1	0.044	-0.277	0.365	
Lettuce	3	4.782	0.246	5.14	5.193	0.352	6.78	0.411	0.309	0.513	0.98
	4	5.527	0.200	3.62	5.638	0.244	4.28	0.111	0.034	0.188	
	5	7.117	0.182	2.56	7.634	0.134	1.76	0.517	0.419	0.615	
	1	3.000	0.155	5.17	3.089	0.158	5.11	0.090	-0.019	0.198	
	2	3.924	0.121	3.08	4.052	0.091	2.25	0.128	0.056	0.199	
Frozen fish	3	4.822	0.174	3.61	5.499	0.139	2.53	0.677	0.518	0.838	0.98
	4	5.860	0.133	2.27	6.246	0.091	1.46	0.386	0.323	0.450	
	5	6.474	0.065	1.00	6.953	0.090	1.29	0.479	0.414	0.544	_
	1	0.881	0.792	89.9	1.012	0.736	72.7	0.130	-0.190	0.450	
	2	2.105	0.154	7.32	2.232	0.157	7.03	0.127	0.026	0.277	
Milk powder	3	4.027	0.133	3.30	3.993	0.076	1.90	-0.034	-0.103	0.035	0.99
	4	5.010	0.110	2.20	4.926	0.103	2.09	-0.084	-0.149	-0.018	
	5	5.720	0.131	2.29	5.639	0.171	3.03	-0.081	-0.142	-0.019	

^oMean of five replicate portions, plated in duplicate, after logarithmic transformation: Log₁₀[CFU/g + (0.1)f].

^bRepeatability standard deviation.

^cRelative standard deviation for repeatability.

^{*d*}Mean difference between the candidate and reference methods.

^eConfidence interval.

^f95% Lower confidence limit for difference of means.

⁹95% Upper confidence limit for difference of means.

^hSquare of correlation coefficient.

		Co	mpact Dry TC 48	า	Co	mpact Dry TC 72	2 h		95%	ί Cl ^e	
Matrix	Cont. level —	Mean ^a	Sr ^b	RSD _r ^c	Mean	S _r	RSDr	Mean diff. ^d	LCL ^f	UCL ^g	– r ^{2h}
	1	0.281	0.594	211	0.281	0.594	211	0.00	0.00	0.00	
	2	2.399	0.304	12.7	2.734	0.636	23.4	0.336	-0.079	0.750	
Cooked chicken	3	3.309	0.364	11.0	3.323	0.369	11.1	0.014	-0.001	0.029	0.99
	4	5.056	0.284	5.62	5.063	0.280	5.53	0.007	0.002	0.011	
	5	6.173	0.252	4.08	6.179	0.249	4.03	0.006	0.003	0.009	
	1	2.346	0.663	28.3	2.496	0.593	23.76	0.149	0.065	0.233	
	2	3.359	0.709	21.1	3.393	0.732	21.6	0.034	0.007	0.061	
Lettuce	3	4.491	0.324	7.21	4.782	0.246	5.14	0.291	0.200	0.381	0.99
	4	5.443	0.201	3.69	5.527	0.200	3.62	0.085	0.001	0.169	
	5	7.010	0.143	2.04	7.117	0.182	2.56	0.107	0.031	0.183	
	1	2.953	0.178	6.03	3.000	0.155	5.17	0.046	0.026	0.067	
	2	3.924	0.121	3.08	3.924	0.121	3.08	0.000	-0.001	0.000	
Frozen fish	3	4.791	0.191	3.99	4.822	0.174	3.61	0.031	0.000	0.063	1.00
	4	5.854	0.135	2.31	5.860	0.133	2.27	0.006	0.002	0.010	
	5	6.432	0.066	1.03	6.474	0.065	1.00	0.043	-0.022	0.107	
	1	0.645	0.845	131	0.881	0.792	89.9	0.236	-0.123	0.596	
	2	2.064	0.139	6.73	2.105	0.154	7.32	0.041	0.008	0.074	
Milk powder	3	4.026	0.134	3.33	4.027	0.133	3.30	0.001	0.000	0.003	0.99
	4	5.010	0.110	2.20	5.010	0.110	2.20	0.000	0.000	0.001	
	5	5.709	0.132	2.31	5.720	0.131	2.29	0.011	0.002	0.020	

^aMean of five replicate portions, plated in duplicate, after logarithmic transformation: Log₁₀[CFU/g + (0.1)f].

^bRepeatability standard deviation.

^cRelative standard deviation for repeatability.

^dMean difference between the candidate and reference methods.

^eConfidence interval.

^f95% Lower confidence limit for difference of means.

^{*g*}95% Upper confidence limit for difference of means.

^{*h*}Square of correlation coefficient.

Table 4. Multi-laboratory study: Summary of pasteurized milk results by laboratory for each contamination level for each method (4)

Laboratory	ι	Uncontaminated	I		Low level			Medium level			High level	
	CD TC ^a 48 h	CD TC 72 h	ISO 4833	CD TC 48 h	CD TC 72 h	ISO 4833	CD TC 48 h	CD TC 72 h	ISO 4833	CD TC 48 h	CD TC 72 h	ISO 4833
1	0.000 ^b	0.000	0.734	2.557	2.560	2.698	3.533	3.538	3.697	4.701	4.703	4.723
2	0.000	0.000	0.000	2.638	2.638	2.832	3.726	3.726	3.825	4.727	4.727	4.860
3	0.521	0.591	0.000	2.684	2.689	2.806	3.763	3.771	3.973	4.803	4.808	4.947
4	2.340	2.365	2.192	2.930	2.937	2.992	3.872	3.909	3.943	4.829	4.852	5.049
5	0.000	0.000	0.000	2.599	2.599	2.762	3.728	3.740	3.805	4.740	4.748	4.814
6	0.000	0.000	0.000	2.795	2.810	2.694	3.737	3.766	3.758	4.811	4.816	4.832
7	0.000	0.000	0.000	2.749	2.749	2.852	3.831	3.831	3.953	4.823	4.825	4.945
8	0.000	0.000	0.000	2.588	2.588	2.770	3.627	3.627	3.917	4.761	4.761	4.826
9	0.260	0.260	0.000	2.751	2.751	2.803	3.790	3.790	3.888	4.904	4.904	4.963
10	0.000	0.260	0.000	2.618	2.618	2.788	3.688	3.688	3.724	4.710	4.710	4.616
11 ^c	0.000	0.000	0.000	2.703	2.705	2.752	3.726	3.726	3.820	4.793	4.793	4.833
12 ^c	0.000	0.000	0.000	2.748	2.753	2.730	3.812	3.734	3.854	4.983	4.940	4.979
13 ^c	0.000	0.000	0.000	2.779	2.779	2.825	3.876	3.878	3.896	4.853	4.858	4.864
14 ^{<i>d</i>}	0.260	0.260	0.000	2.700	2.700	2.810	3.739	3.739	3.789	4.835	4.836	4.833
14 ^{<i>c,d</i>}	0.260	0.260	0.000	2.700	2.700	2.745	3.774	3.774	3.833	4.716	4.716	4.701

^aCompact Dry TC.

^bResults are reported for each laboratory as a mean of 2 replicate portions, plated in duplicate, after logarithmic transformation: Log₁₀[CFU/g + (0.1)f].

^cSample analysis initiated one day later than scheduled start date.

^dOrganizing Laboratory.

Table 6. Multi-la	boratory study: I	Method cor	nparison of	pasteurized	l milk resul	lts for each	contaminatio	on level acr	oss 14 data	sets ^a (4)								
		Compact D	ory TC 48 h v	rs ISO 4833				Compact D	Dry TC 72 h v	s ISO 4833				Con	npact Dry TC	48 h vs 72 h		
Contam. level	Mean [♭] CD TC 48 h	Mean ISO 4833	Mean diff. ^c	LCL ^d	UCL ^e	r ^{2f}	Mean CD TC 72 h	Mean ISO 4833	Mean diff.	LCL	UCL	r²	Mean CD TC 48 h	Mean CD TC 72 h	Mean diff.	LCL	UCL	r²
Uncont.	0.112	0.052	-0.060	-0.270	0.080		0.117	0.052	-0.065	-0.281	0.079		0.112	0.117	0.005	-0.006	0.018	
Low	2.686	2.776	0.090	0.046	0.129	0.98	2.688	2.776	0.088	0.043	0.129	0.98	2.686	2.688	0.002	-0.003	0.006	1.0
Mid	3.737	3.838	0.101	0.064	0.131	0.98	3.738	3.838	0.100	0.065	0.134	0.98	3.737	3.738	0.001	-0.011	0.008	1.0
High	4.797	4.838	0.041	0.011	0.069		4.796	4.838	0.042	0.012	0.070		4.797	4.796	-0.001	-0.008	0.006	

^aLaboratory 4 removed from statistical analysis due to unexpectedly high counts in the uncontaminated samples. See Table 4.

^bOverall mean across laboratories for each method, after logarithmic transformation: Log₁₀[CFU/g + (0.1)f].

^cMean difference between methods.

^{*d*}95% Lower confidence limit for difference of means.

^e95% Upper confidence limit for difference of means.

^fSquare of correlation coefficient.

DISCUSSION OF MODIFICATION STUDY APPROVED DECEMBER 2020 (5)

The results of this study indicate that the CompactDry "Nissui" TC method can be used for rapid and accurate enumeration of mesophilic aerobic bacteria in raw ground beef and raw chicken breast at 24 h of incubation. The CompactDry "Nissui" TC method showed similar repeatability to the MLG 3.02 reference method and equivalent mean results. Enumeration was equivalent between 24 h and 48 h for the CompactDry "Nissui" TC.

The CompactDry "Nissui" TC method offers a time saving of 1 day over the reference method and produces a mesophilic aerobic bacterial count in 24 ± 2 h. Advantages of the CompactDry "Nissui" TC method include reductions in storage space, media preparation, incubator space, and waste disposal.

able 4. Method compa	arison data	a sumr	mary and sta	atistics for 24	h incubation of	ⁱ candidate me	thod (5)							
			Compa	actDry "Nissu	i" TC, 24 h		MLG 3.02			95 %	% CI [∌]	90 9	% Cl ^b	
			Mean			Mean								
	Cont.		Log ₁₀			Log ₁₀								
Matrix	level	Ν	CFU/g	Sr	RSD _r , %	CFU/g	Sr	RSD _r , %	DOM ^a	LCL ^c	UCL^d	LCL ^c	UCL ^d	Equivalence
Raw ground beef,	Low	5	2.189	0.120	5.48	2.368	0.109	4.60	-0.180	-0.389	0.030	-0.340	-0.019	Passed
95% lean	Med	5	5.519	0.214	3.88	5.881	0.177	3.01	-0.362	-0.495	-0.229	-0.464	-0.260	Passed
(naturally	High	5	7.399	0.174	2.35	7.577	0.068	0.90	-0.177	-0.357	0.002	-0.315	-0.040	Passed
contaminated)														
Raw chicken breast,	Low	5	3.458	0.061	1.76	3.390	0.035	1.03	0.068	-0.041	0.177	-0.016	0.151	Passed
98% lean	Med	5	5.691	0.447	7.85	5.788	0.303	5.23	-0.097	-0.350	0.156	-0.292	0.097	Passed
(naturally contaminated)	High	5	7.487	0.334	4.46	7.333	0.260	3.55	0.153	-0.018	0.324	0.022	0.284	Passed

^aDOM = Difference of Means

^bCl = Confidence Interval for DOM

^cLCL = Lower confidence limit for DOM

^dUCL = Upper confidence limit for DOM

able 5. Method compa	arison data	a sumr	nary and sta	tistics for 48	h incubation of o	andidate meth	rod (5)							
			CompactDry "Nissui" TC, 48 h			MLG 3.02				95 % Cl ^b		90 % Cl ^b		
			Mean			Mean							_	
	Cont.		Log ₁₀			Log ₁₀								
Matrix	level	Ν	CFU/g	Sr	RSD _r , %	CFU/g	Sr	RSD _r , %	DOM ^a	LCL ^c	UCL ^d	LCL ^c	UCL ^d	Equivalence
Raw ground beef,	Low	5	2.552	0.145	5.68	2.368	0.109	4.60	0.184	0.016	0.352	0.055	0.313	Passed
95% lean	Med	5	5.694	0.202	3.55	5.881	0.177	3.01	-0.186	-0.310	-0.063	-0.281	-0.091	Passed
(naturally	High	5	7.478	0.122	1.63	7.577	0.068	0.90	-0.099	-0.215	0.018	-0.188	-0.009	Passed
contaminated)														
Raw chicken breast,	Low	5	3.685	0.065	1.76	3.390	0.035	1.03	0.295	0.199	0.391	0.222	0.369	Passed
98% lean	Med	5	5.728	0.457	7.98	5.788	0.303	5.23	-0.060	-0.343	0.222	-0.277	0.156	Passed
(naturally	High	5	7.518	0.319	4.24	7.333	0.260	3.54	0.185	0.013	0.356	0.053	0.317	Passed
contaminated)														

^{*a*}DOM = Difference of Means

^bCI = Confidence Interval for DOM

^cLCL = Lower confidence limit for DOM

^dUCL = Upper confidence limit for DOM

Table 6. Method compa	arison data	a sumr	nary and sta	atistics for 24	h vs. 48 h incuba	tion of candida	ate method	(5)						
Matrix	Cont. level		CompactDry "Nissui" TC, 24 h			CompactDry "Nissui" TC, 48 h				95 % CI [♭]		90 % Cl ^b		
		N	Mean		RSDr, %	Mean			DOMa					Equivalence
			Log10 CFU/g	Sr		Log₁₀ CFU/g	Sr	RSD _r , %		LCL ^c	UCL ^d	LCL ^c	UCL ^d	
Raw ground beef,	Low	5	2.189	0.120	5.48	2.552	0.145	5.68	-0.363	-0.465	-0.262	-0.441	-0.286	Passed
95% lean	Med	5	5.519	0.214	3.88	5.694	0.202	3.55	-0.176	-0.206	-0.146	-0.199	-0.153	Passed
(naturally contaminated)	High	5	7.399	0.174	2.35	7.478	0.122	1.63	-0.079	-0.157	0.000	-0.139	-0.019	Passed
Raw chicken breast,	Low	5	3.458	0.061	1.76	3.685	0.065	1.76	-0.228	-0.295	-0.161	-0.279	-0.176	Passed
98% lean	Med	5	5.691	0.447	7.85	5.728	0.457	7.98	-0.037	-0.081	0.007	-0.070	-0.003	Passed
(naturally contaminated)	High	5	7.487	0.334	4.46	7.518	0.319	4.24	-0.032	-0.083	0.020	-0.071	0.008	Passed

^aDOM = Difference of Means

^bCI = Confidence Interval for DOM

^cLCL = Lower confidence limit for DOM

 d UCL = Upper confidence limit for DOM

REFERENCES CITED

- 1. Kodaka, H., Mizuochi, S., Teramura, H., and Nirazuka, T., Comparison of the Compact Dry TC Method with the Standard Pour Plate Method (AOAC Official Method 966.23) for Determining Aerobic Colony Counts in Food Samples, AOAC *Performance Tested Methods*SM certification number 010404.
- 2. Official Methods of Analysis 19th Ed., AOAC INTERNATIONAL, Rockville, MD, Method 966.23, Microbiological Methods, http://www.eoma.aoac.org, accessed 2004
- 3. ISO 4833:2003, Microbiology of food and animal feeding stuffs Horizontal method for the enumeration of microorganisms Colony-count technique at 30°C, http://www.iso.org/iso/iso_catalogue_tc/catalogue_tc/catalogue_detail.htm?csnumber=34524, accessed 2006
- 4. Mizuochi, S., Evaluation of Compact Dry TC: Matrix Extension, AOAC Performance Tested MethodsSM certification number 010404. Modification approved April 2015.
- 5. Toyota, K., Modification of the CompactDry "Nissui" TC for Enumeration of Aerobic Bacteria at 24 Hours in raw Ground Beef and Raw Chicken Breast, AOAC *Performance Tested Methods*SM certification number 010404. Approved December 2020.
- 6. FSIS MLG 3.02 Quantitative Analysis of Bacteria in Foods as Sanitary Indicators: Chapter 3.6 Aerobic Plate Count https://www.fsis.usda.gov/wps/wcm/connect/f2162091-af72-4888-997b-718d6592bcc9/MLG-3.pdf?MOD=AJPERES accessed July 17, 2020